

# Discovery of a Potent, $S1P_3$ -Sparing Benzothiazole Agonist of Sphingosine-1-Phosphate Receptor 1 ( $S1P_1$ )

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**ABSTRACT** Optimization of a benzofuranyl S1P<sub>1</sub> agonist lead compound (3) led to the discovery of 1-(3-fluoro-4-(5-(2-fluorobenzyl)benzo[d]thiazol-2-yl)benzyl)azetidine-3-carboxylic acid (14), a potent S1P<sub>1</sub> agonist with minimal activity at S1P<sub>3</sub>. Dosed orally at 0.3 mg/kg, 14 significantly reduced blood lymphocyte counts 24 h postdose and attenuated a delayed type hypersensitivity (DTH) response to antigen challenge.

**KEYWORDS** Sphingosine-1-phosphate receptor, S1P<sub>1</sub>, agonist, inflammation, lymphocyte

Research efforts over the past decade have revealed the lysophospholipid sphingosine-1-phosphate (1, S1P; Figure 1) to be a pleiotropic modulator of diverse cellular processes including migration, adhesion, proliferation, and differentiation.<sup>1,2</sup> These effects are mediated, at least in part, by the interaction of S1P with a set of paralogous G protein-coupled receptors (S1P<sub>1-5</sub>), which are widely expressed in the immune, cardiovascular, and central nervous systems.<sup>3-5</sup> The S1P<sub>1</sub> receptor, highly expressed in lymphocytes, has recently been the focus of extensive investigation due to its role in the regulation of lymphocyte egress from lymph nodes.<sup>6-9</sup>

Much of this effort has centered on the study of fingolimod (2, FTY-720),<sup>10</sup> a synthetic amino diol prodrug, which is stereoselectively phosphorylated in vivo by sphingosine kinase 2 to generate the corresponding (*S*)-phosphate (FTY-720P), which is a potent agonist of  $S1P_{1,3-5}$ . FTY-720P binding to lymphocyte  $S1P_1$  receptors leads to receptor internalization (RI) and proteolysis.<sup>11</sup> Knockout studies in mice have revealed  $S1P_1$  receptor expression in  $T^{12,13}$  and B cells<sup>14</sup> to be required for these lymphocytes to exit secondary lymphoid organs and enter the circulation; FTY-720P-mediated RI thus leads to lymphocyte sequestration in secondary lymphoid organs.<sup>15</sup>

Although FTY-720-mediated lymphocyte sequestration does not impair antiviral and antibacterial immune responses to secondary infections in rodent models,<sup>16,17</sup> decreased lymphocyte recruitment to sites of inflammation has been demonstrated to moderate immune responses in rodent models of multiple sclerosis<sup>18,19</sup> and organ transplantation.<sup>20</sup> Phase III human clinical trials have recently revealed **2** to be efficacious in reducing relapse rates in patients with relapsing—remitting multiple sclerosis.<sup>21,22</sup> Human clinical trials have additionally shown dosing of **2** to be associated with transient, dose-dependent decreases in mean heart rate and asymptomatic atrioventricular blockade.<sup>23</sup> Studies of **2** in rodents have revealed lymphocyte sequestration and heart rate reduction to be driven by distinct S1P receptor isoforms (S1P<sub>1</sub> and S1P<sub>3</sub>, respectively), however.<sup>24–27</sup> These observations, coupled with the impressive efficacy of **2** emerging from human clinical investigations, have prompted the wide-spread search for S1P<sub>1</sub> agonists with minimal activity at the S1P<sub>3</sub> receptor.<sup>28,29</sup> In this communication, we describe the discovery of a potent S1P<sub>3</sub>-sparing S1P<sub>1</sub> agonist, benzothiazole **14**, as well as its in vivo pharmacological and pharmacokinetic characteristics.

In prior work, in silico docking of a library of commercially available molecules with druglike properties into a S1P<sub>1</sub> receptor model generated using PREDICT methodology,<sup>30</sup> followed by modeling-driven structure—activity relationship (SAR) studies, led to the identification of a benzofuran-containing hit series, of which **3** was chosen as the lead molecule.<sup>31</sup> Benzofuran **3** demonstrated double-digit nano-molar activity at hS1P<sub>1</sub> in an assay measuring RI of an hS1P<sub>1</sub>-GFP fusion protein in U2OS cells and limited activity at hS1P<sub>3</sub> as determined by Ca<sup>2+</sup> mobilization in hS1P<sub>3</sub>- and G<sub>q/i5</sub>- transfected CHO-K1 cells (Table 1). Oral dosing (1 mg/kg) of **3** in rats resulted in a statistically significant reduction in circulating lymphocytes 24 h postdose and significantly

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Figure 1. S1P, FTY-720, and benzofuran lead 3.

 Table 1. SAR of 5-Benzyl-Substituted Bicyclic Cores<sup>a</sup>



compd	W	Х	Y	Z	$hS1P_1 RI$ $EC_{50} (\mu M)$ (% efficacy)	$\begin{array}{c} hS1P_3 \ Ca^{2+} \\ EC_{50} \ (\mu M) \\ (\% \ efficacy) \end{array}$
3	0	С	СН	С	0.057 (96)	1.73 (42)
4	Ο	С	Ν	С	0.325(105)	0.354 (77)
5	Ν	С	S	С	0.221 (84)	3.47 (50)
6	Ν	С	NH	С	> 6.25	> 25
7	Ν	С	CH	Ν	6.72 (115)	> 25
8	Ν	Ν	СН	С	2.89(53)	1.11 (43)

<sup>*a*</sup> Data represent an average of at least two determinations. Percent efficacy is reported relative to  $1.00 \,\mu$ M **9** and  $0.200 \,\mu$ M S1P for hS1P<sub>1</sub> RI and hS1P<sub>3</sub> Ca<sup>2+</sup> assays, respectively. (S1P demonstrates 91% efficacy relative to  $1.00 \,\mu$ M **9** in hS1P<sub>1</sub> RI studies.) > [Highest concentration tested] is reported for compounds that do not achieve > 10% of control activity ( $1.00 \,\mu$ M **9** or  $0.200 \,\mu$ M S1P).

attenuated a delayed type hypersensitivity (DTH) response to antigen challenge.<sup>32</sup> One potential liability of **3** identified during preclinical toxicology studies was the observation of proconvulsive activity in rat at oral doses of 40 mg/kg and higher. Herein, we report on a successful effort to identify derivatives of **3** lacking this liability.

As proconvulsive activity had not been reported with prior nonisoform-selective S1P receptor agonists, such activity was tentatively assumed to be peculiar to benzofuran **3**. An effort was therefore undertaken to identify closely structurally related analogues of **3** that possessed similar or improved pharmacological profiles but that were devoid of proconvulsive activity. To this end, a series of compounds was prepared wherein the benzofuran core of **3** was replaced with a range of bioisosteric heterocycles (Tables 1 and 2). Analogues wherein the benzylic substituent of the central 5,6-ring system was located para to the more electronegative substituent ("W") of the six-membered ring were uniformly found to be less potent (4–118-fold) than **3** in the S1P<sub>1</sub> RI assay (Table 1). In contrast, several analogues wherein the benzylic substituent was located meta to the pubs.acs.org/acsmedchemlett

 Table 2.
 SAR of 6-Benzyl-Substituted Bicyclic Cores<sup>a</sup>



compd	W	Х	Y	Z	$hS1P_1$ RI EC <sub>50</sub> ( $\mu$ M) (% efficacy)	$\begin{array}{c} hS1P_{3}\\ Ca^{2+} EC_{50} (\mu M)\\ (\% \text{ efficacy})\end{array}$
9	0	С	CH	С	0.013 (110)	0.679(42)
10	0	С	Ν	С	0.094 (108)	0.542 (87)
11	Ν	С	S	С	0.042(102)	1.21 (24)
12	Ν	С	CH	Ν	0.741 (110)	6.09 (50)
13	Ν	Ν	CH	С	1.81 (100)	2.71 (48)

 $^{\it a}$  Data represent an average of at least two determinations. Percent efficacy is reported as in Table 1.

 Table 3. Optimization of Benzothiazole Agonists<sup>a</sup>



compd	R	hS1P <sub>1</sub> RI EC <sub>50</sub> ( $\mu$ M) (% efficacy)	hS1P <sub>3</sub> Ca <sup>2+</sup> EC <sub>50</sub> (μM) (% efficacy)
11	Н	0.042(102)	1.21 (24)
14	2-F	0.042(115)	3.47(14)
15	3-F	0.021 (138)	0.687(22)
16	4-F	0.033 (129)	1.91 (37)
17	2,4-di-F	0.042(186)	1.20 (24)
18	2,6-di-F	0.126(126)	> 25
19	3,4-di-F	0.030(151)	0.967(19)
20	2-Me	0.026(118)	2.44(40)
21	3-Me	0.054(92)	> 2.5
22	4-Me	0.077(132)	1.80 (17)
23	2-Cl	0.042(121)	4.88(41)
24	3-Cl	0.049(141)	3.88(35)
25	4-Cl	0.159(117)	> 25

<sup>*a*</sup> Data represent an average of at least two determinations. Percent efficacy is reported as in Table 1. >[Highest concentration tested] is reported for compounds that do not achieve > 10% of control activity (1.00  $\mu$ M **9** or 0.200  $\mu$ M S1P).

more electronegative substituent of the six-membered ring possessed comparable or superior  $S1P_1$  activity to **3** (Table 2). Benzothiazole **11** was particularly noteworthy, as it possessed not only comparable in vitro activity to lead **3** but proved similarly active in vivo, bringing about statistically significant lymphocyte depletion upon oral dosing at 1.0 mg/kg.

In an effort to further understand the  $S1P_1$  and  $S1P_3$  SARs of benzothiazole **11**, a series of analogues was prepared wherein the terminal phenyl group of **11** was modified with a range of small lipophilic substituents (Table 3).  $S1P_1$  activity was generally maintained across these analogues; however,

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**Figure 2.** Compound 14 dosed orally reduces blood lymphocyte counts in female Lewis rats 24 h postdose (N = 5/group; bars represent average blood lymphocyte counts + SEs; circles represent average plasma concentration  $\pm$  SEs; \*P < 0.05 vs vehicle by ANOVA/Dunnett's multiple comparison test; vehicle = 20 % captisol, pH 2).

4-substitution of the phenyl group with larger substituents led to moderate decreases in S1P<sub>1</sub> activity (**22**, EC<sub>50</sub> = 0.077  $\mu$ M; **25**, EC<sub>50</sub> = 0.159  $\mu$ M), as did 2,6-difluorination (**18**, EC<sub>50</sub> = 0.126  $\mu$ M). Substitution of the terminal phenyl group demonstrated a more varied effect on S1P<sub>3</sub> activity: Whereas methylation had little impact on S1P<sub>3</sub> activity (EC<sub>50</sub> = 1.80–2.44  $\mu$ M), chlorination led to moderate to significant decreases in S1P<sub>3</sub> activity (EC<sub>50</sub> = 3.88 to >25  $\mu$ M), and fluorination resulted both in compounds with increased S1P<sub>3</sub> activity (**15**, EC<sub>50</sub> = 0.687  $\mu$ M) and in a compound with no detectable activity at S1P<sub>3</sub> (**18**, EC<sub>50</sub> > 25  $\mu$ M).

One trend to emerge from these studies was that 2-substitution of the terminal phenyl group of **11** uniformly led to moderate, but significant, reductions in S1P<sub>3</sub> activity. As a small lipophilic substituent proved nearly equally effective to a larger substituent in reducing S1P<sub>3</sub> activity (cf. **14**, CLog P = 4.1, and **23**, CLog P = 4.6),<sup>33</sup> we selected the least lipophilic analogue, 2-fluorobenzyl benzothiazole **14** (RI EC<sub>50</sub> =  $0.042 \,\mu$ M, Ca<sup>2+</sup> EC<sub>50</sub> =  $3.47 \,\mu$ M), for further in vitro and in vivo profiling.<sup>34</sup>

Dosed orally at 0.3, 1.0, and 3.0 mg/kg in Lewis rats, **14** produced a dose-dependent reduction in circulating blood lymphocytes 24 h postdose, consistent with S1P<sub>1</sub> agonism (Figure 2). Statistical significance (P < 0.05) was achieved at a dose of 0.3 mg/kg (34% reduction in lymphocytes vs vehicle), and a 3.0 mg/kg dose resulted in near maximal lymphopenia (74% reduction in lymphocytes vs vehicle).<sup>35</sup>

Compound 14 was subsequently investigated in a DTH antigen challenge model, an in vivo model of cell-mediated immune responses. In this study, Lewis rats were orally dosed with 0.1, 0.3, 1.0, or 3.0 mg/kg 14 or vehicle once daily over the course of 10 days. One day after initiating dosing, rats were immunized with a mixture of ovalbumin and complete Freund's adjuvant. One week later, the immunized rats were rechallenged with ovalbumin by intracutaneous injection into one ear. After an additional 2 days, differential swelling of the inoculated ear (determined by change in ear thickness) and circulating blood lymphocyte counts were measured. As seen in Figure 3, a statistically significant (P < 0.05) reduction in ear swelling was observed at doses of 0.3 mg/kg and higher. Furthermore, reduced ear swelling was found to closely track



Figure 3. Compound 14 dosed orally (qd for 10 days) reduces ovalbumin (OVA)-induced ear swelling in OVA-immunized female Lewis rats 48 h post-OVA challenge [N = 8/group (vehicle, 0.1, 0.3, and 1.0 mg/kg), N = 4/group (3.0 mg/kg); bars represent average change in ear thickness + SEs; circles represent average blood lymphocyte counts  $\pm$  SEs; \*P < 0.05 vs vehicle by ANOVA/Dunnett's multiple comparison test; vehicle = 20% captisol, pH 2].

Table 4. Pharmacokinetic Parameters for 14

species	Cl (L/h/kg)	V <sub>ss</sub> (L/kg)	$T_{1/2}(h)$	MRT (h)	% F
rat <sup>a</sup>	0.33	3.3	7.5	10	68
$\mathrm{NHP}^{b}$	0.50	1.6	35.2	3.3	23

<sup>*a*</sup> Female Sprague–Dawley (iv: 2 mg/kg, DMSO, N = 3; po: 15 mg/kg, 20% captisol, pH 2, N = 3). <sup>*b*</sup> Male Cynomolgus (iv: 4 mg/kg, 20% captisol, pH 4, N = 3; po: 10 mg/kg, 20% captisol, 1% pluronic F68, 1% HPMC, pH 2, N = 3).

circulating lymphocyte counts, consistent with the cellmediated etiology of the DTH immune response.

Pharmacokinetic profiling of 14 in rats and nonhuman primates (Table 4) revealed 14 to possess acceptable characteristics for further development: 14 demonstrated low clearance (10–19% of liver blood flow), moderate steady state volumes of distribution ( $V_{ss} = 1.6-3.3 \text{ L/kg}$ ), moderate-to-long mean residence times (3.3–10 h), and acceptable oral bioavailability (23–68%). In vitro studies established that 14 neither inhibited nor induced human cytochrome P450 enzymes, was nonmutagenic (Ames and micronucleus negative), and did not significantly inhibit the hERG channel in an electrophysiology assay (PatchXpress;<sup>36</sup> IC<sub>50</sub> > 10  $\mu$ M).

Cardiovascular safety studies of **14** in telemetered rats established a no-effect level for heart rate and mean arterial pressure changes of 30 mg/kg (po), indicating a wide margin for S1P<sub>3</sub>-associated cardiovascular toxicity. Most significantly, **14** did not exhibit proconvulsive activity in Sprague–Dawley rats (N = 3) at a dose of 100 mg/kg when administered orally once daily for 4 days. The exposure of **14** at the end of the fourth day ( $C_{max} = 49 \pm 10 \mu g/mL$ , AUC<sub>0-24</sub> = 987 ± 185  $\mu g \cdot h/mL$ ) was significantly higher (3-fold  $C_{max}$ , 40-fold AUC<sub>0-24</sub>) than the exposure at which proconvulsive activity was observed with benzofuran **3**. As the plasma free fractions<sup>37</sup> of **14** (2.1%) and **3** (1.5%) in rat were similar, the lack of proconvulsive activity of **14** relative to **3** is currently not well understood.

The synthesis of benzothiazole **14** is described in Scheme 1. Aryl bromide **26**, prepared in one step from commercially available 3-fluoro-4-bromobenzaldehyde, was treated with Scheme 1. Synthesis of 14<sup>*a*</sup>



<sup>*a*</sup> Reagents and conditions: (a) *n*-BuLi, **27**, THF, -78 °C, 63%. (b) Na<sub>2</sub>CO<sub>5</sub>, DMF, 110 °C, 93%. (c) 5 N HCl, THF, 55 °C, 100%. (d) Methyl azetidine-3-carboxylate hydrochloride, AcOH, DIPEA, NaBH<sub>3</sub>CN, MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 71%. (e) Bis(di-*tert*-butyl(phenyl)phosphine) palladium dichloride (5 mol %), (2-fluorobenzyl)zinc chloride, THF, 85%. (f) LiOH, THF/H<sub>2</sub>O, then HCl and pH 6 sodium phosphate buffer, 99%.

*n*-butyllithium at -78 °C, and 2-fluoro-5-bromophenyl isothiocyanate **27** was then added to the resulting aryl lithium species, generating the corresponding thioamide. Incubation of the thioamide with a suspension of sodium carbonate in hot DMF resulted in clean cyclization to benzothiazole **28**. Exposure of **28** to HCl in warm THF resulted in cleavage of the acetal protecting group; reductive amination of the resulting aldehyde subsequently provided methyl azetidine carboxylate **29**. Negishi coupling of **29** with (2-fluorobenzyl)zinc chloride delivered 5-benzylbenzothiazole **30**, and saponification of **30** followed by pH adjustment of the resulting reaction mixture resulted in the precipitation of benzothiazole **14** as its zwitterion.

In summary, replacement of the benzofuranyl core of a novel  $S1P_1$  agonist (3) with a series of bioisosteric heterocycles led to the identification of benzothiazole 11, which retained both the S1P<sub>1</sub> activity and the S1P<sub>3</sub> selectivity of  $\mathbf{3}$ . Subsequent substitution of the terminal phenyl group of 11 with a range of small lipophilic substituents led to the discovery of 2-fluorobenzyl benzothiazole 14, which was found to be a potent  $S1P_1$  agonist (EC<sub>50</sub> = 0.042  $\mu$ M) with reduced activity at  $S1P_3$ (EC<sub>50</sub> = 3.47  $\mu$ M). Oral dosing of 14 in rats (0.3 mg/kg) resulted in a statistically significant reduction in circulating lymphocytes 24 h postdose, as well as a statistically significant attenuation of a DTH response to antigen challenge. Furthermore, benzothiazole 14 was devoid of the proconvulsive activity of 3 at doses of up to 100 mg/kg. In vitro and in vivo pharmacokinetic studies and safety studies have established that 14 possesses acceptable properties for further development.

**SUPPORTING INFORMATION AVAILABLE** Statistical analysis of data presented in Tables 1–3, experimental procedures and characterization data for **3–25**, and details of in vitro and in vivo assays. This material is available free of charge via the Internet at http://pubs.acs.org.

### AUTHOR INFORMATION

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### REFERENCES

- Hannun, Y. A.; Obeid, L. M. Principles of Bioactive Lipid Signaling: Lessons from Sphingolipids. *Nature Rev. Mol. Cell. Biol.* 2008, 9, 139–150.
- (2) Kihara, A.; Mitsutake, S.; Mizutani, Y.; Igarashi, Y. Metabolism and Biological Functions of Two Phosphorylated Sphingolipids, Sphingosine-1-Phosphate and Ceramide 1-Phosphate. *Prog. Lipid Res.* 2007, 46, 126–144.
- (3) Rosen, H.; Gonzalez-Cabrera, P. J.; Sanna, M. G.; Brown, S. Sphingosine 1-Phosphate Receptor Signaling. *Annu. Rev. Biochem.* 2009, 78, 743–768.
- Cyster, J. G. Chemokines, Spingosine-1-Phosphate, and Cell Migration in Secondary Lymphoid Organs. *Annu. Rev. Immunol.* 2005, 23, 127–159.
- (5) Rosen, H.; Goetzl, E. J. Sphingosine 1-Phosphate and its Receptors: An Autocrine and Paracrine Network. *Nature Rev. Immunol.* 2005, *5*, 560–570.
- (6) Marsolais, D.; Rosen, H. Chemical Modulators of Sphingosine-1-Phosphate Receptors as Barrier-Oriented Therapeutic Molecules. *Nature Rev. Drug Discovery* **2009**, *8*, 297–307.
- (7) Brinkmann, V. Sphingosine 1-Phosphate Receptors in Health and Disease: Mechanistic Insights from Gene Deletion Studies and Reverse Pharmacology. *Pharmacol. Ther.* 2007, *115*, 84–105.
- (8) Gardell, S. E.; Dubin, A. E.; Chun, J. Emerging Medicinal Roles for Lysophospholipid Signaling. *Trends Mol. Med.* 2006, *12*, 65–75.
- Mandala, S.; Hajdu, R.; Bergstrom, J.; Quackenbush, E.; Xie, J.; Milligan, J.; Thornton, R.; Shei, G.-J.; Card, D.; Keohane, C.; Rosenbach, M.; Hale, J.; Lynch, C. L.; Rupprecht, K.; Parsons, W.; Rosen, H. Alteration of Lymphocyte Trafficking by Sphingosine-1-Phosphate Receptor Agonists. *Science* 2002, *296*, 346–349.
- (10) Adachi, K.; Chiba, K. FTY720 Story. Its Discovery and the Following Accelerated Development of Sphingosine 1-Phosphate Receptor Agonists as Immunomodulators Based on Reverse Pharmacology. *Perspect. Med. Chem.* **2007**, *1*, 11–23.
- (11) Oo, M. L.; Thangada, S.; Wu, M.-T.; Liu, C. H.; Macdonald, T. L.; Lynch, K. R.; Lin, C.-Y.; Hla, T. Immunosuppressive and Antiangiogenic Sphingosine 1-Phosphate Receptor-1 Agonists Induce Ubiquitinylation and Proteasomal Degradation of the Receptor. J. Biol. Chem. 2007, 282, 9082–9089.
- (12) Matloubian, M.; Lo, C. G.; Cinamon, G.; Lesneski, M. J.; Xu, Y.; Brinkmann, V.; Allende, M. L.; Proia, R. L.; Cyster, J. G. Lymphocyte Egress from Thymus and Peripheral Lymphoid Organs is Dependent on S1P Receptor 1. *Nature* **2004**, *427*, 355–360.
- (13) Allende, M. L.; Dreier, J. L.; Mandala, S.; Proia, R. L. Expression of the Sphingosine 1-Phosphate Receptor, S1P1, on T-cells Controls Thymic Emigration. *J. Biol. Chem.* 2004, 279, 15396–15401.
- (14) Kabashima, K.; Haynes, N. M.; Xu, Y.; Nutt, S. L.; Allende, M. L.; Proia, R. L.; Cyster, J. G. Plasma Cell S1P1 Expression Determines Secondary Lymphoid Organ Retention Versus Bone Marrow Tropism. *J. Exp. Med.* **2006**, *203*, 2683–2690.
- (15) Chiba, K.; Yanagawa, Y.; Masubuchi, Y.; Kataoka, H.; Kawaguchi, T.; Ohtsuki, M.; Hoshino, Y. FTY720, a Novel Immunosuppressant, Induces Sequestration of Circulating Mature Lymphocytes

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by Acceleration of Lymphocyte Homing in Rats. I. FTY720 Selectively Decreases the Number of Circulating Mature Lymphocytes by Acceleration of Lymphocyte Homing. *J. Immunol.* **1998**, *160*, 5037–5044.

- (16) Pinschewer, D. D.; Ochsenbein, A. F.; Odermatt, B.; Brinkmann, V.; Hengartner, H.; Zinkernagel, R. M. FTY720 Immunosuppression Impairs Effector T Cell Peripheral Homing Without Affecting Induction, Expansion, and Memory. *J. Immunol.* 2000, *164*, 5761–5770.
- (17) Kursar, M.; Jänner, N.; Pfeffer, K.; Brinkmann, V.; Kaufmann, S. H. E.; Mittrücker, H.-W. Requirement of Secondary Lymphoid Tissues for the Induction of Primary and Secondary T cell Responses Against *Listeria monocytogenes. Eur. J. Immunol.* **2008**, *38*, 127–138.
- (18) Kataoka, H.; Sugahara, K.; Shimano, K.; Teshima, K.; Koyama, M.; Fukunari, A.; Chiba, K. FTY720, Sphingosine 1-Phosphate Receptor Modulator, Ameliorates Experimental Autoimmune Encephalomyelitis by Inhibition of T Cell Infiltration. *Cell. Mol. Immunol.* 2005, *2*, 439–448.
- (19) Fujino, M.; Funeshima, N.; Kitazawa, Y.; Kimura, H.; Amemiya, H.; Suzuki, S.; Li, X.-K. Amelioration of Experimental Autoimmune Encephalomyelitis in Lewis Rats by FTY720 Treatment. *J. Pharmacol. Exp. Ther.* **2003**, *305*, 70–77.
- (20) Chiba, K.; Hoshino, Y.; Suzuki, C.; Masubuchi, Y.; Yanagawa, Y.; Ohtsuki, M.; Sasaki, S.; Fujita, T. FTY720, A Novel Immunosuppressant Possessing Unique Mechanisms. I. Prolongation of Skin Allograft Survival and Synergistic Effect in Combination with Cyclosporine in Rats. *Transplant. Proc.* **1996**, *28*, 1056–1059.
- (21) Kappos, L.; Radue, E.-W.; O'Connor, P.; Polman, C.; Hohlfeld, R.; Calabresi, P; Selmaj, K.; Agoropoulou, C.; Leyk, M.; Zhang-Auberson, L.; Burtin, P. A Placebo-Controlled Trial of Oral Fingolimod in Relapsing Multiple Sclerosis. *N. Engl. J. Med.* **2010**, *362*, 387–401.
- (22) Cohen, J. A.; Barkhof, F.; Comi, G.; Hartung, H.-P.; Khatri, B. O.; Montalban, X.; Pelletier, J.; Capra, R.; Gallo, P.; Izquierdo, G.; Tiel-Wilck, K.; de Vera, A.; Jin, J.; Stites, T.; Wu, S.; Aradhye, S.; Kappos, L. Oral Fingolimod or Intramuscular Interferon for Relapsing Multiple Sclerosis. *N. Engl. J. Med.* **2010**, *362*, 402–415.
- (23) See refs 21, 22, and Schmouder, R.; Serra, D.; Wang, Y.; Kovarik, J. M.; DiMarco, J.; Hunt, T. L.; Bastien, M.-C. FTY720: Placebo-Controlled Study of the Effect on Cardiac Rate and Rhythm in Healthy Subjects. *J. Clin. Pharmacol.* **2006**, *46*, 895–904.
- (24) Forrest, M.; Sun, S.-Y.; Hajdu, R.; Bergstrom, J.; Card, D.; Doherty, G.; Hale, J.; Keohane, C.; Meyers, C.; Milligan, J.; Mills, S.; Nomura, N.; Rosen, H.; Rosenbach, M.; Shei, G.-J.; Singer, I. I.; Tian, M.; West, S.; White, V.; Xie, J.; Proia, R. L.; Mandala, S. Immune Cell Regulation and Cardiovascular Effects of Sphingosine-1-Phosphate Receptor Agonists in Rodents are Mediated via Distinct Receptor Subtypes. *J. Pharmacol. Exp. Ther.* **2004**, *309*, 758–768.
- (25) Sanna, M. G.; Liao, J.; Jo, E.; Alfonso, C.; Ahn, M.-Y.; Peterson, M. S.; Webb, B.; Lefebvre, S.; Chun, J.; Gray, N.; Rosen, H. Sphingosine 1-Phosphate (S1P) Receptor Subtypes S1P<sub>1</sub> and S1P<sub>3</sub>, Respectively, Regulate Lymphocyte Recirculation and Heart Rate. *J. Biol. Chem.* **2004**, *279*, 13839–13848.
- (26) Recent research has suggested that  $S1P_1$  rather than  $S1P_3$  mediates heart rate reduction in humans: Gergely, P.; Wallström, E.; Nuesslein-Hildesheim, B.; Bruns, C.; Zécri, F.; Cooke, N.; Traebert, M.; Tuntland, T.; Rosenberg, M.; Saltzman, M. Phase I study with the selective  $S1P_1/S1P_5$  receptor modulator BAF312 indicates that  $S1P_1$  rather than  $S1P_3$  mediates transient heart rate reduction in humans. *Mult. Scler.* **2009**, *15*, S125–S126. See also ref 27.

- (27) Brossard, P.; Hofmann, S.; Cavallaro, M.; Halabi, A.; Dingemanse, J. Entry-into-Humans Study with ACT-128800, A Selective S1P<sub>1</sub> Receptor Agonist: Tolerability, Safety, Pharmacokinetics, and Pharmacodynamics. *Clin. Pharmacol. Ther.* **2009**, *85*, S63–S64.
- (28) Buzard, D. J.; Thatte, J.; Lerner, M.; Edwards, J.; Jones, R. M. Recent Progress in the Development of Selective S1P1 Receptor Agonists for the Treatment of Inflammatory and Autoimmune Disorders. *Expert Opin. Ther. Patents* **2008**, *18*, 1141–1159.
- (29) Cooke, N.; Zécri, F. Sphingosine 1-Phosphate Type 1 Receptor Modulators: Recent Advances and Therapeutic Potential. *Annu. Rep. Med. Chem.* 2007, *42*, 245–263. For reports from 2008 to the present and modeling-based approaches to rationalizing isoform selectivity, see the Supporting Information.
- (30) Shacham, S.; Marantz, Y.; Bar-Haim, S.; Kalid, O.; Warshaviak, D.; Avisar, N.; Inbal, B.; Heifetz, A.; Fichman, M.; Topf, M.; Naor, Z.; Noiman, S.; Becker, O. M. PREDICT Modeling and In-Silico Screening for G-Protein Coupled Receptors. *Proteins* **2004**, *57*, 51–86.
- (31) Saha, A. K.; Yu, X.; Lin, J.; Lobera, M.; Sharadendu, A.; Cheruku, S.; Schutz, N.; Segal, D.; Marantz, Y.; McCauley, D.; Buys, J.; Horner, M.; Salyers, K.; Schrag, M.; Vargas, H. M.; Xu, Y.; McElvain, M.; Xu, H. Benzofuran-based Sphingosine-1 Phosphate Receptor-1 (S1P1) Agonists: Discovery of a Potent, Selective and Orally Bioavailable Preclinical Lead Molecule. *ACS Med. Chem. Lett.*, in press.
- (32) Ovalbumin/complete Freund's adjuvant. For experimental protocol, see the Supporting Information.
- (33) ChemBioDraw Ultra, version 11.0, Cambridgesoft, 100 CambridgePark Drive, Cambridge, MA 02140.
- (34) Dosed orally (1 mg/kg) in female Lewis rats, 14, 20, and 23 were similarly efficacious in reducing blood lymphocyte counts 24 h postdose (63-73% reduction).
- (35) S1P<sub>1</sub> agonists have previously been reported to produce a maximal ~70% reduction in lymphocyte count: Hale, J. J.; Lynch, C. L.; Neway, W.; Mills, S. G.; Hajdu, R.; Keohane, C. A.; Rosenbach, M. J.; Milligan, J. A.; Shei, G.-J.; Parent, S. A.; Chrebet, G.; Bergstrom, J.; Card, D.; Ferrer, M.; Hodder, P.; Strulovici, B.; Rosen, H.; Mandala, S. A Rational Utilization of High-Throughput Screening Affords Selective, Orally Bioavailable 1-Benzyl-3-carboxyazetidine Sphingosine-1-phosphate-1 Receptor Agonists. *J. Med. Chem.* 2004, *47*, 6662– 6665.
- (36) PatchXpress 7000A Automated Parallel Patch-Clamp System, Molecular Devices, 1311 Orleans Drive, Sunnyvale, CA 94089.
- (37) Determined by ultracentrifugation at 5.0  $\mu$ g/mL test compound concentration.